

Feasibility of a new fully automated ADAMTS13 activity assay

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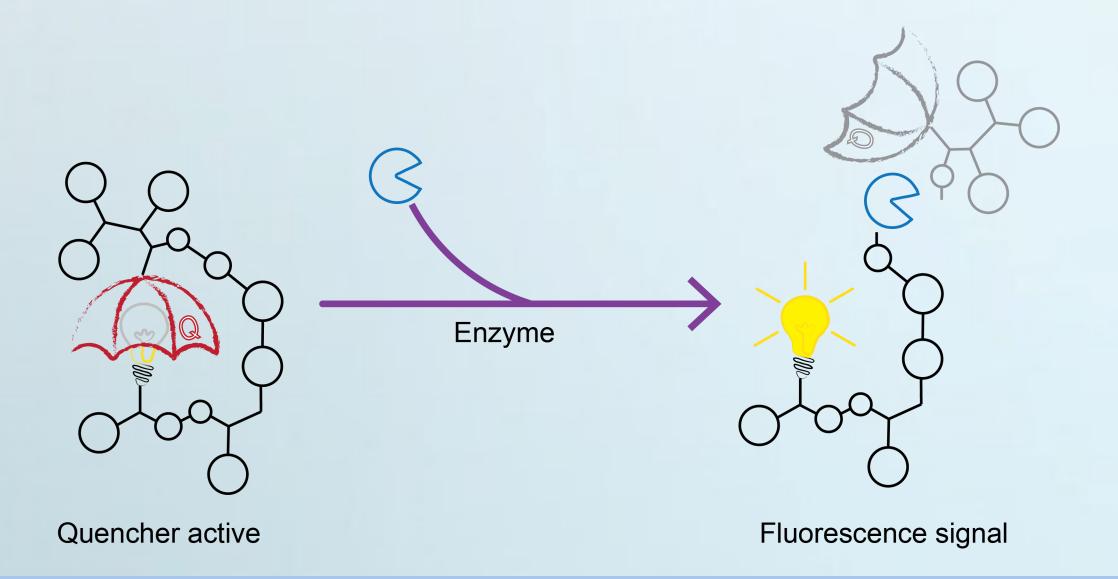
INTRODUCTION

The aim of these studies was to develop a fully automated ADAMTS13 activity assay using a fluorescence resonance energy transfer (FRET) method. For this purpose a new coagulation analyzer, equipped with an optical Quenching module was co-developed. Generally, the new ADAMTS13 activity assays should benefit from an automated analytical process (eliminating errors, improve quality, reduce work load, turn-around time and costs) while maintaining the key performance parameters of the well established TECHNOZYM ADAMTS-13 Activity ELISA (LOQ, linearity) to ensue patient safety

MATERIALS AND METHODS

Assay principle

The newly developed TECHNOFLUOR ADAMTS13 Activity employs a FRET substrate based on the VWF73. Upon cleavage by sample derived ADAMTS13, the quenching part is removed and flourescent signal is emmitted. The measurable signal is proportional to the ADAMTS13 activity level.





A newly developed automated analyzer with Quenching module (Ceveron s100), combining routine and speciality haemostasis testing, was used.

RESULTS

Calibration

With an assay time of <30 min a calibration curve ranging from 0 - 0.8 IU/mL could be established exhibiting excellent recovery of controls being calibrated with TECHNOZYM ELISA system.

High Control (0.71 IU/mL) - 101% recovery

Low Control (0.16 IU/mL) - 94% recovery

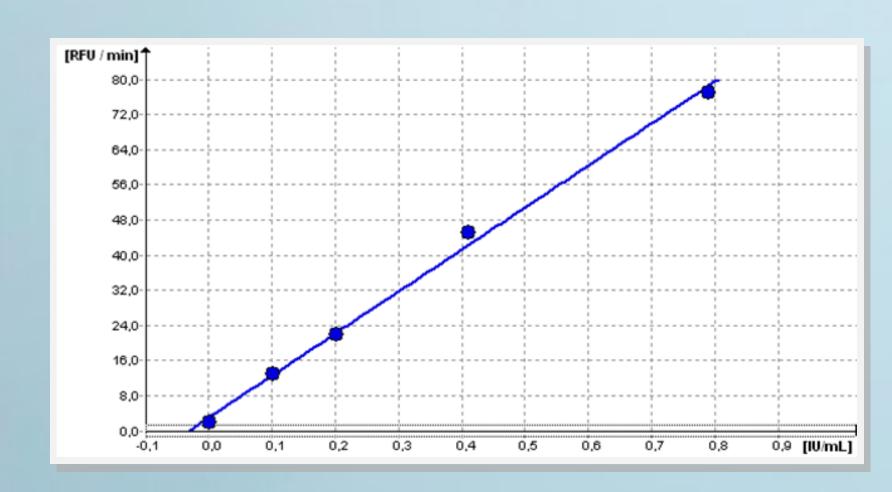


Fig 1. typical calibration curve on Ceveron s100 traceable to the WHO standard

Method comparison

The linear correlation (Passing and Bablok regression) between TECHNOZYM ELISA results and the new automated method (TECHNOFLUOR) shows a good correlation coefficient (r>0.90) and an intercept of <0.01 IU/mL.

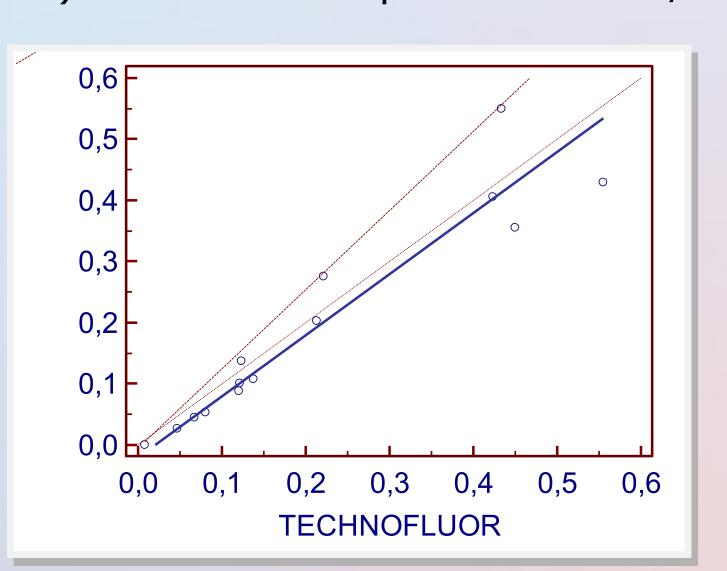


Fig 2. Method comparison to TECHNOZYM ADAMTS13 Activity ELISA

Stability

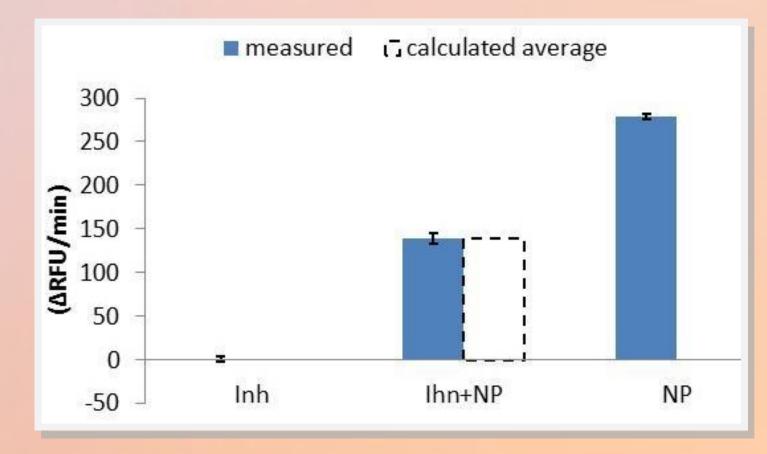
To investigate the stability of the new ADAMTS13 activity substrate three samples with different activity levels were tested. Lyophilized substrate was compared fresh and stored for 1 month at 37°C.

	ADAMTS13 Activity	Recovery
Sample 1	0.9 IU/mL	96.5 %
Sample 2	0.4 IU/mL	105.8 %
Sample 3	0.1 IU/mL	93.5 %

Effect of inhibitors

Analyzing an automated mixing study for ADAMTS13 we studied an

EQA sample from ECAT (18 INH units). Although the measured activity was <0.1 IU/mL, the sample mixed with normal pooled plasma (1+1) did not show effects of the ADAMTS13 inhibitors.



CONCLUSIONS

The fully automated ADAMTS13 Activity test run on the new Ceveron s100 haemostasis analyzer is a fast, easy to use and accurate option to improve ADAMTS13 testing for faster diagnostics and better treatment of TMA patients. With a high degree of linearity over the whole assay range, the new TECHNOFLUOR ADAMTS13 Activity assay shows excellent comparison to both reference methods for ADAMTS13 (FRETS and chromogenic ELISA).

Based on the stability experiments we expect a minimum shelf life of 24 months. The use of lyophilized INH samples might be problematic (known incompatibility for Bethesda-like test systems) and further studies need to test fresh or frozen samples.